

Note

Polysaccharides from the roots of the marsh mallow (*Althaea officinalis* L., var. *Rhobusta*): dianhydrides of oligosaccharides of the aldose type

PETER CAPEK, DUŠAN UHRÍN, JOZEF ROSÍK, ALŽBETA KARDOŠOVÁ, RUDOLF TOMAN, AND VINCENT MIHÁLOV

Institute of Chemistry, Centre for Chemical Research, Slovak Academy of Sciences, 842 38 Bratislava (Czechoslovakia)

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Partial hydrolysis¹ of the heteropolysaccharide complex from the roots of the marsh mallow (*Althaea officinalis* L., var. *Rhobusta*) (a medicinal plant), which is composed of D-galacturonic acid, L-rhamnose, D-glucuronic acid, and D-galactose in the molar ratios 1:1:1:1.2, afforded, in addition to reducing oligosaccharides, two other oligosaccharides (**1** and **2**). Oligosaccharide **1** had $[\alpha]_D + 76.5^\circ$ (c 1, water) and was composed of D-galacturonic acid and L-rhamnose; oligosaccharide **2**, which had $[\alpha]_D + 51.6^\circ$ (c 1, water), contained D-galacturonic acid, L-rhamnose, and D-glucuronic acid in equimolar ratios. These compounds did not reduce Fehling's solution or picric acid and were unaffected by sodium borohydride, indicating that all of the anomeric carbons were involved in linkages.

A non-reducing oligosaccharide containing D-galacturonic acid and L-rhamnose has been described^{2,3}, which was shown³ to be a 1,2':2,1'-dianhydride of α -D-galactopyranuronic acid and β -L-rhamnopyranose. According to our knowledge, a non-reducing oligosaccharide containing D-galacturonic acid, L-rhamnose, and D-glucuronic acid has not been described hitherto. We now report elucidation of the structures of **1** and **2**, using ¹H- and ¹³C-n.m.r. spectroscopy.

The ¹H-n.m.r. data of **1** and **2** are listed in Table I. Comparison of the ¹H-n.m.r. data of **1** with those for L-rhamnopyranose⁴ and D-galactopyranuronic⁵ acid revealed similar coupling constants. The $J_{1',2'}$ value of 3.7 Hz (cf. 3.0 and 7.5 Hz for α - and β -D-galacturonic acid⁵, respectively) indicated the D-galacturonic acid residue to be α . The $J_{1,2}$ value of 1.3 Hz for the L-rhamnose residue indicated⁴ it to be β . In

TABLE I

¹H-N.M.R. DATA FOR 1 AND 2

Compound	Unit	Chemical shifts (δ, p.p.m.)						Coupling constants (Hz)					
		H-1	H-2	H-3	H-4	H-5	H-6	J _{1,2}	J _{2,3}	J _{3,4}	J _{4,5}	J _{5,6}	
1	D-GalA	5.29	3.92	4.78	4.39	4.77		3.7	10.4	3.4	1.4		
	L-Rha	4.89	4.25	3.77	3.49 ^a	3.38 ^a	1.36 ^a	1.3	3.8	9.6 ^a	9.4 ^a		4.0 ^a
2	D-GlcA	5.03	3.37 ^b	3.56 ^b	3.61 ^b	3.97		7.9	^c	^c	9.5 ^a		
	D-GalA	5.28	4.07	4.94	4.54	4.78		3.7	10.3	3.4	1.3		
	L-Rha	4.89	4.24	3.76	3.46 ^b	3.37 ^b	1.37	1.3	3.8	9.6 ^a	^c		4.0 ^a

^aApproximation to first order. ^bAssigned on the basis of a 2D-hetero-correlated experiment. ^cOverlapping multiplets.

spite of non-reducing properties of **1**, the chemical shifts for H-1 of the D-galacturonic acid (5.29 p.p.m.) and L-rhamnose residues (4.89 p.p.m.) differed only slightly from those of H-1 in α -D-galactopyranuronic acid (5.31 p.p.m.) and β -L-rhamnopyranose (4.86 p.p.m.). However, the H-3' signal was shifted markedly to low field (4.78 p.p.m.) in comparison with that (3.84 p.p.m.) of H-3 for α -D-galacturonic acid⁵, and H-2 of the L-rhamnose residue was deshielded (0.31 p.p.m.) compared to the corresponding proton in L-rhamnose. Fujiwara and Arai³ observed a similar large deshielding of H-3' in the 1,2':2,1'-dianhydride of 3,4-di-O-acetyl- β -L-rhamnopyranose and methyl 3,4-di-O-acetyl- α -D-galactopyranuronate compared to that in methyl 3,4-di-O-acetyl-2-O-(methyl 2,3,4-tri-O-acetyl- α -D-galactopyranosyluronate)- β -L-rhamnopyranoside. Therefore, the deshielding of H-3 is indicative of the presence of a 1,4-dioxane grouping between the D-galacturonic acid and L-rhamnopyranose residues in **1**. Confirmation of the presence of two glycosidic bonds between the two units in **1** was provided by ¹³C-n.m.r. spectroscopy. Starting from the known ¹H chemical shifts of **1**, the ¹³C chemical shifts (Table II) were assigned by the result of a 2D-heterocorrelated experiment⁶. The presence of two pairs of heteronuclear three-bond couplings through glycosidic bonds was then established. Using the selective transfer of polarisation⁷ from H-1, H-2', H-1', and H-2, the signals of C-2', C-1, C-2, and C-1', respectively, appeared in the individual spectra (some other carbon signals of the respective saccharide units were also observed). These experiments confirmed the existence of (1 \rightarrow 2')- and (1' \rightarrow 2)-glycosidic linkages in **1**.

The 1,4-dioxane grouping in **1** significantly affected the chemical shifts of the resonances for C-1 and C-2 in each unit. These nuclei were shielded (> 5 p.p.m.) in comparison to the respective carbons in β -L-Rha-(1 \rightarrow 3)- α -D-Gal⁸ and β -D-Gal-(1 \rightarrow 2)- β -L-Rha⁹ (reference compounds for the L-rhamnose moiety), and α -D-Gal-(1 \rightarrow 2)- α -D-ManOR¹⁰ and α -L-Fuc-(1 \rightarrow 2)- α -D-Gal¹¹ (reference compounds for the D-galacturonic acid moiety). Thus, **1** is identified as α -D-galactopyranuronic acid β -L-rhamnopyranose 1,2':2,1'-dianhydride.

The ¹H-n.m.r. data (Table I) for the D-galacturonic acid and L-rhamnose

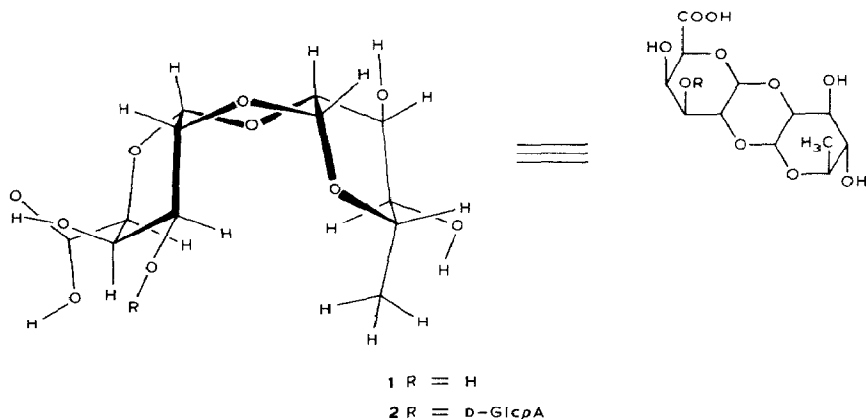


TABLE II

¹³C-N.M.R. CHEMICAL SHIFTS^a FOR **1** AND **2**

Compound	Unit	Chemical shifts (δ, p.p.m.)					
		C-1	C-2	C-3	C-4	C-5	C-6
1	D-GalA	95.9	71.6	69.7	71.5	73.9	173.2
	L-Rha	92.6	77.0	72.1	72.8	73.2	18.2
2	D-GlcA	103.8	74.0	76.3	72.4	75.6	174.4
	D-GalA	95.6	70.6	77.3	71.1	73.3	172.9
	L-Rha	92.4	76.8	72.0	72.6	73.1	18.3

^aFor solutions in D₂O (internal MeOH).

residues in **2** are similar to those in **1**, indicating the presence of two glycosidic linkages between the D-galacturonic acid and L-rhamnose residues.

In the region for anomeric protons, there was a signal at 5.03 p.p.m. (*J* 7.9 Hz) consistent with a non-reducing β-D-glucopyranuronic acid residue¹². The ¹³C signals for **2** were assigned by the results of a 2D-heterocorrelated experiment⁶. Based on the known ¹H chemical shifts, most ¹³C resonances were assigned unambiguously. The signals of C-2'', C-3'', and C-4'' (of D-glucuronic acid) and C-4 and C-5 (of L-rhamnose) could not be assigned in this way due to overlapping of the respective ¹H signals. The assignment of these resonances was aided by comparison with chemical shifts of the ¹³C resonances for methyl β-D-glucopyranosiduronic acid¹³ and **1**. The ¹³C chemical shifts (Table II) for the signals of the corresponding carbon atoms of the D-galacturonic acid and L-rhamnose residues of **1** and **2** were similar except for that of the resonance of C-3' of the D-galacturonic acid residue (**1**, 69.7 p.p.m.; **2**, 77.3 p.p.m.), reflecting 3'-substitution in **2**. The chemical shifts of the remaining signals for **2** were similar to those of the corresponding carbon reso-

TABLE III

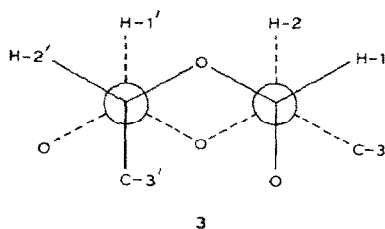
³J_{H,C} VALUES FOR **1**^a

Coupling constant	Value (Hz)	Dihedral angle ^d (degrees)
<i>J</i> _{H-2,C-1}	6.2 ^b	-170
<i>J</i> _{H-1,C-2'}	1.8 ^c	+55
<i>J</i> _{H-1',C-2}	6.2 ^b	+170
<i>J</i> _{H-2',C-1}	2.5 ^c	-50

^aAccurate to ±0.1 Hz. ^bBy selective 2D INEPT¹⁶. ^cBy a selective 2D *J*-resolved experiment¹⁷.^dDetermined from the Karplus-type dependence¹⁵; negative sign represents anticlockwise rotation.

nances for methyl β -D-glucopyranosiduronic acid¹³. Considering the similarity of the ^1H - and ^{13}C -n.m.r. data for the D-galacturonic acid and L-rhamnose residues in **2** and **1**, and the data for the D-glucuronic acid residue, **2** is identified as 3-O-(β -D-glucopyranosyluronic acid)- α -D-galactopyranuronic acid β -L-rhamnopyranose 1,2':2,1'-dianhydride.

The stereochemistry of the central dioxane ring in **1** and **2** was determined on the basis of the appropriate $^3J_{\text{H,C}}$ values, which show Karplus-type dependence^{14,15} on dihedral angle and were measured in a 2D J -resolved experiment^{16,17}. The results obtained are summarised in Table III. The dihedral angles found correspond to a chair conformation (**3**) of the central 1,4-dioxane ring. The resulting 1C_4 and 4C_1 conformations of the L-rhamnopyranose and D-galacturonic acid residues, respectively, accord with those found by Fujiwara and Arai³.



The dianhydride **1** has been isolated also from an acid hydrolysate of Karaya gum² and by derivatisation of 2-O-(α -D-galactopyranosyluronic acid)-L-rhamnopyranose³. The formation of **1** and **2**, as well as of dianhydrides of higher oligosaccharides, occurred¹ on acid hydrolysis of a heteropolysaccharide complex, the main chain of which is composed of (1 \rightarrow 4)-linked D-galacturonic acid and (1 \rightarrow 2)-linked L-rhamnose residues. It appears that the intermediate C-1 carbocation of L-rhamnose, formed on splitting of the (1 \rightarrow 2) linkage between L-rhamnose and D-galacturonic acid, may approach and react with HO-2 of the D-galacturonic acid due to rotation around the C-1-C-2 bond, so that a 1,4-dioxane ring is formed between the saccharide units, the chair conformation of which is determined by the 4C_1 and 1C_4 conformations of D-galacturonic acid and L-rhamnose, respectively.

The formation of dianhydrides of the aldose type under conditions of acid hydrolysis is being studied further.

EXPERIMENTAL

Oligosaccharides **1** and **2** were obtained from a partial acid hydrolysate of the heteropolysaccharide complex¹, applying the separation method described by Larsson².

N.m.r. spectra were recorded on 4% solutions in D_2O (^1H , internal DSS; ^{13}C , internal MeOH) with a Bruker AM-300 spectrometer.

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